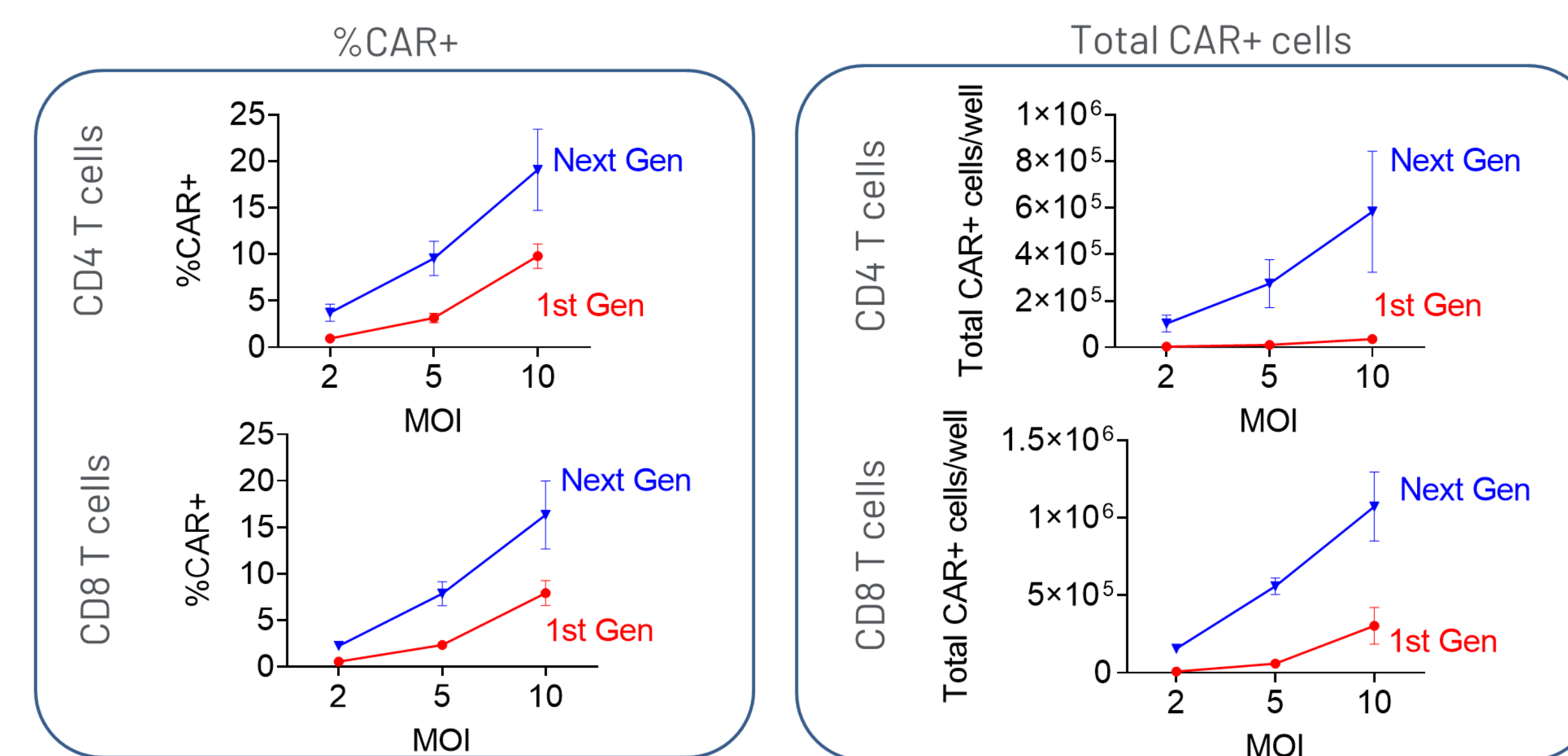


## Abstract

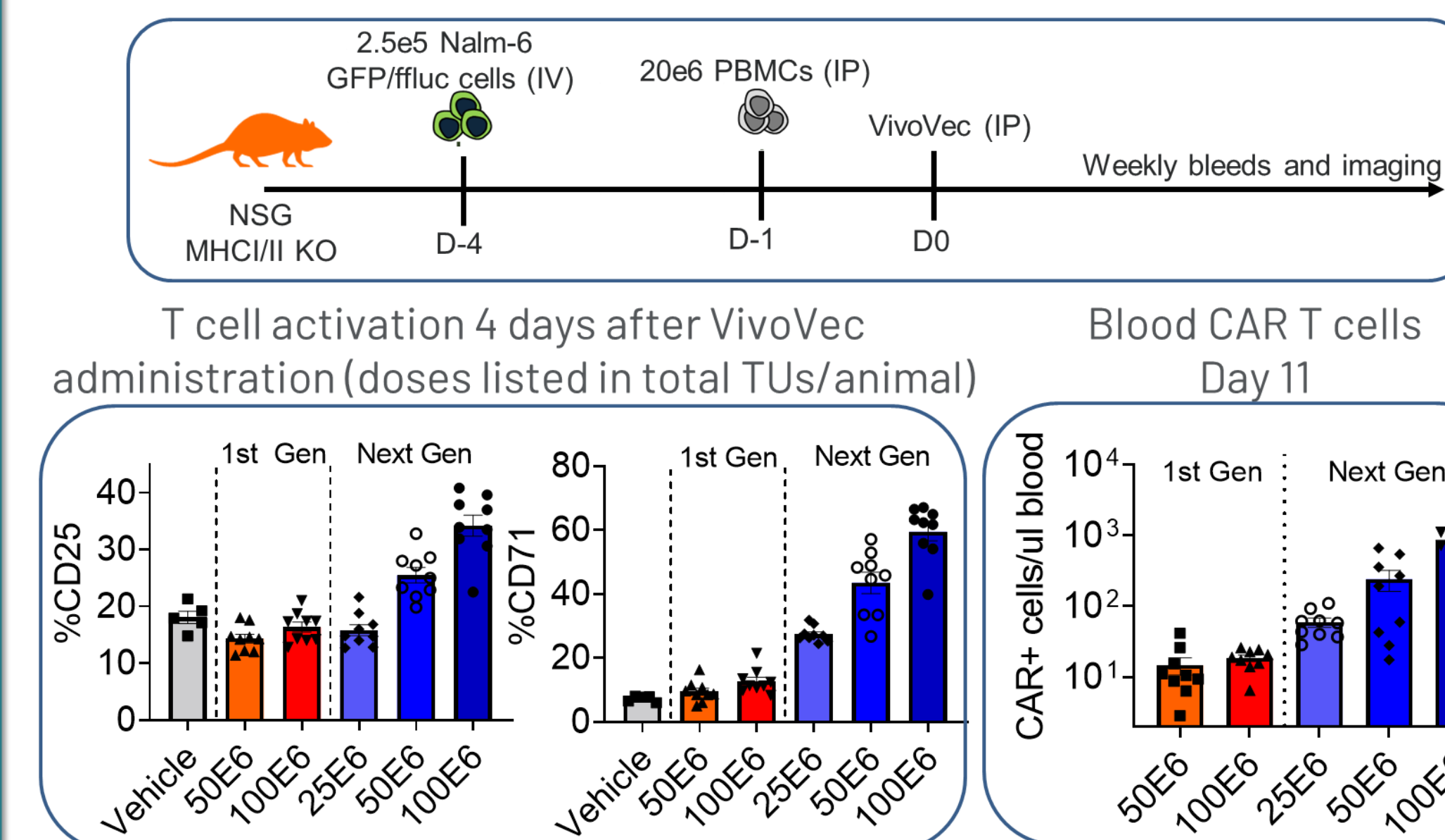
Autologous chimeric antigen receptor (CAR) T cell therapies have revolutionized the treatment of B cell malignancies, leading to long-term remission in 30–40% of certain patient populations. Despite the promising clinical efficacy of CAR T cells in hematologic malignancies, major limitations hinder their widespread application, including challenges for patient access, complex manufacturing, and high cost. To overcome these challenges, we have developed VivoVec, a surface-engineered lentiviral vector-based platform harboring a CAR transgene that is being developed for off-the-shelf use for the generation of CAR T cells *in vivo*. To achieve specific and efficient *in vivo* T cell transduction, VivoVec particles are pseudotyped with the Cocal fusion glycoprotein and an anti-CD3 single chain variable fragment (scFv), and we have previously shown that these first-generation particles generate CAR T cells *in vivo* that mediate antitumor activity. We have advanced the VivoVec platform through incorporating costimulatory molecules into the particle surface, in addition to the anti-CD3 scFv and Cocal fusion glycoprotein. These next generation VivoVec particles exhibit enhanced T cell binding and activation, resulting in increased transduction and greater numbers of CAR<sup>+</sup> T cells *in vitro*. In addition, CAR T cells generated with next generation VivoVec particles exhibited a less-differentiated, central memory-like phenotype and enhanced CAR-antigen-specific polyfunctionality, including cytokine production, proliferation, and tumor cell killing. Finally, in a humanized NSG mouse model of B cell malignancy we observed that next generation VivoVec particles generated greater numbers of CAR T cells in the blood, resulting in enhanced antitumor activity at lower doses compared to first-generation particles. Our results indicate that incorporation of costimulatory molecules onto the surface of VivoVec particles increases both the overall number and functionality of the resulting CAR T cells, greatly augmenting VivoVec mediated CAR T cell generation and antitumor activity *in vivo*. Overall, these data demonstrate that next generation VivoVec particles efficiently generate large numbers of highly functional CAR T cells able to mediate durable tumor control in a preclinical model of B cell malignancy. VivoVec particles have the potential to overcome many of the limitations associated with the current class of CAR T cell therapies.

## Fig 2. Next Gen Particles incorporating costimulatory molecules increase transduction frequency and T cell expansion *in vitro*

VivoVec particles packaging an anti-CD19 CAR were added to PBMCs from 3 donors. 7 days later % of T cells (**Left**) and total number of CAR<sup>+</sup> T cells/well (**Right**) were analyzed.

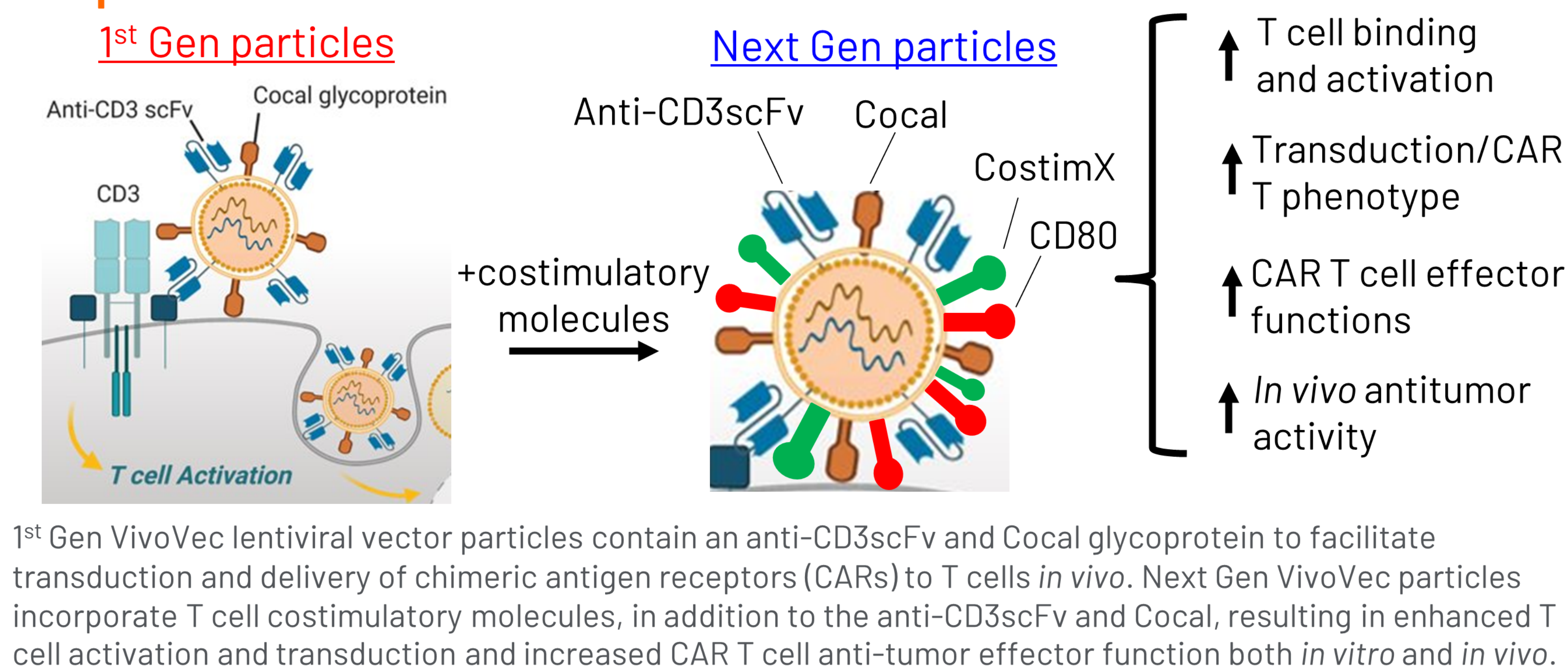


## Fig 5. Next Gen VivoVec particles drive dose-dependent T cell activation and transduction *in vivo* in a tumor xenograft model

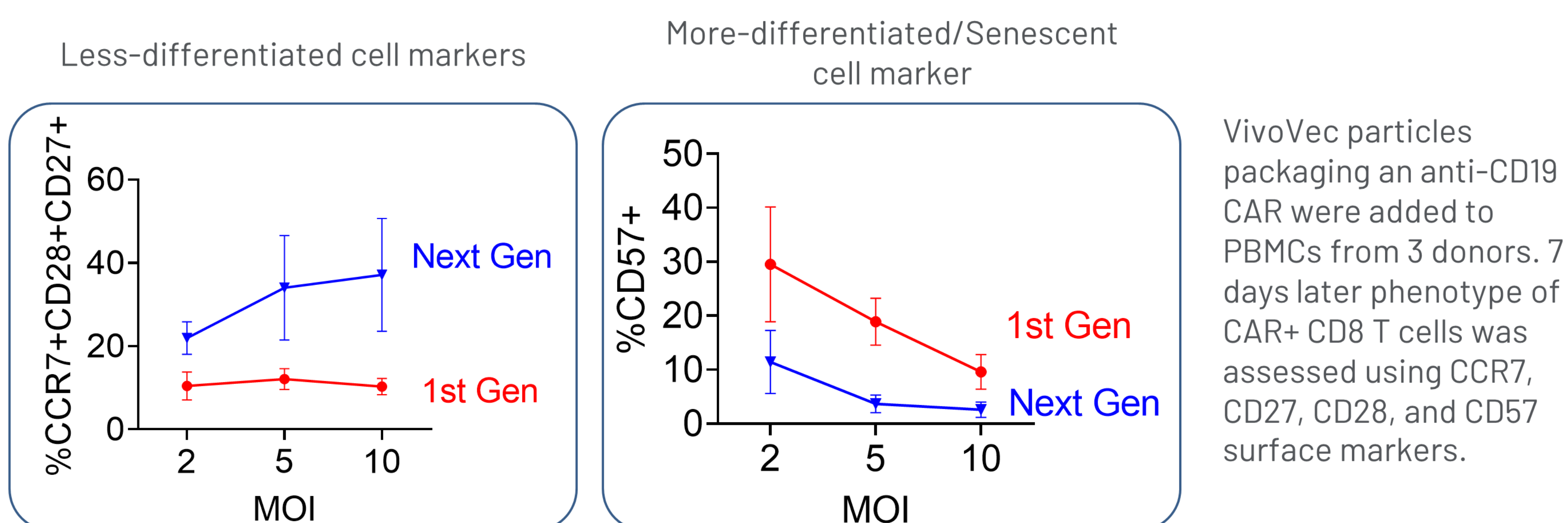


**Top:** Study outline. Luciferase+ Nalm6 Tumor-bearing NSG MHCII KO mice were humanized by injecting 20e6 PBMCs into the intraperitoneal cavity. Mice were then given varying doses of VivoVec lentiviral vector particles packaging an anti-CD19 CAR transgene. **Left:** Activation makers on CD3<sup>+</sup> T cells in the blood 4 days after VivoVec administration. Doses in TUs (Transducing Units). **Right:** Total anti-CD19 CAR<sup>+</sup> CD3<sup>+</sup> T cells found in the blood on Day 11.

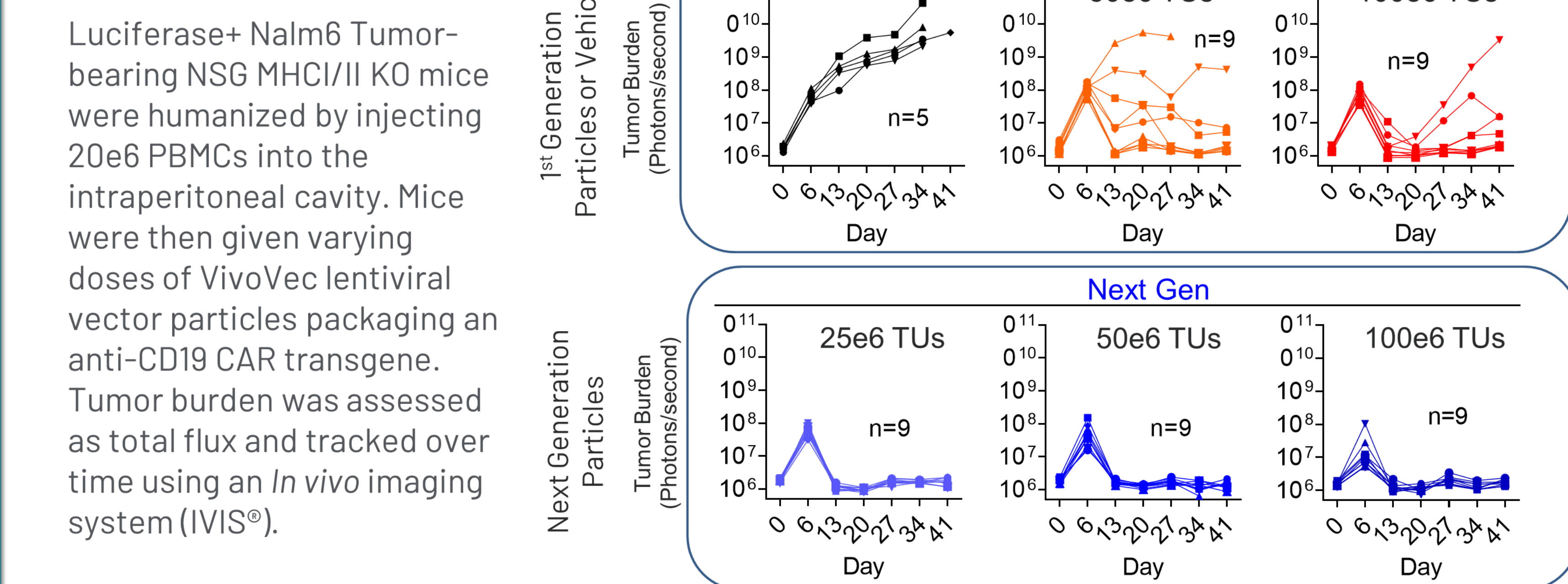
## Graphical Abstract



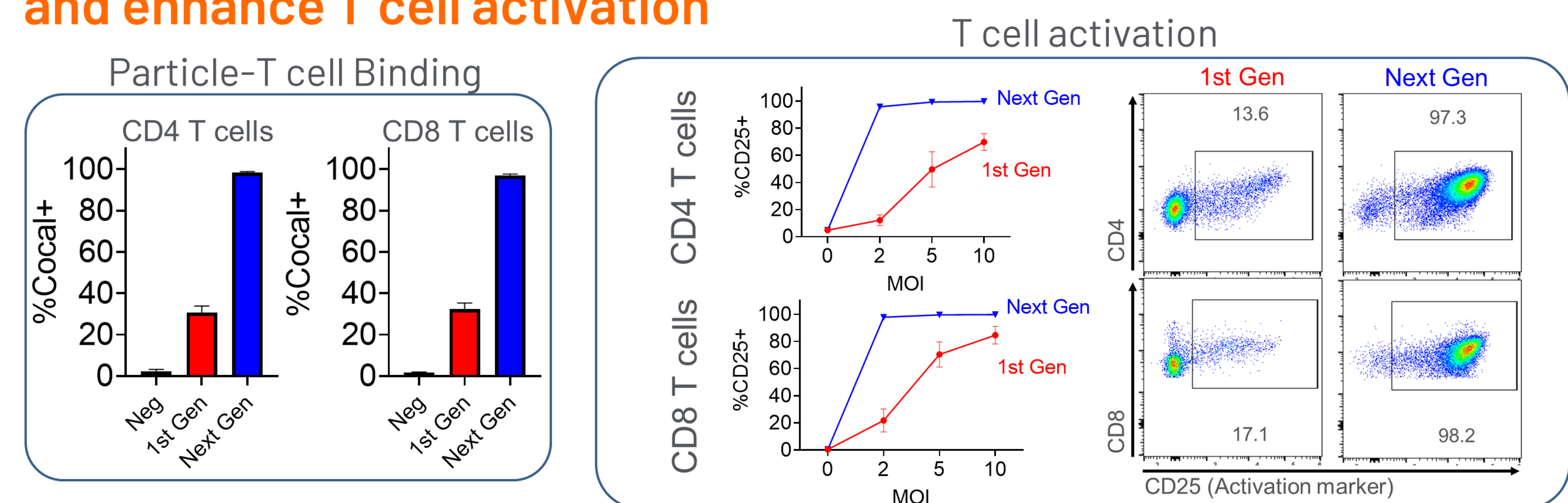
## Fig 3. CAR T cells generated with Next Gen Surface-engineered particles display a less differentiated phenotype



## Fig 6. Next Gen VivoVec particles demonstrate enhanced tumor control *in vivo*

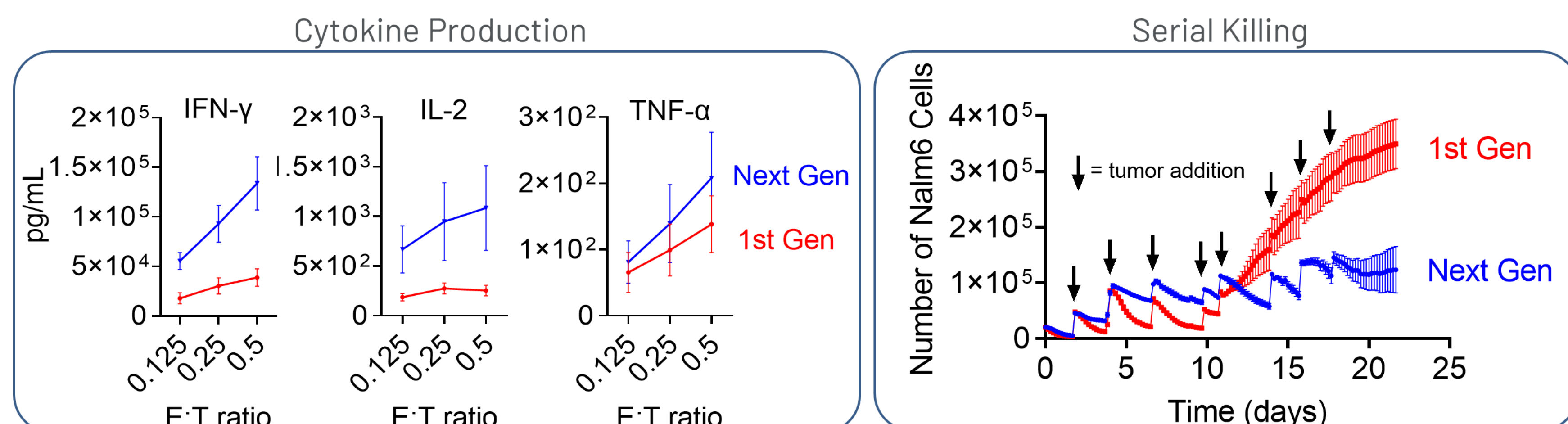


## Fig 1. Next Gen particles demonstrate increased binding to T cells and enhance T cell activation



**Left:** Vector Binding Assay. Particles cultured with PBMCs for 6 hours followed by surface staining for Cocal glycoprotein on T cells. 3 PBMC donors. **Right:** CD25 expression on T cells 3 days after transduction with VivoVec particles. 3 PBMC donors. Flow plots taken from representative samples at MOI=2.

## Fig 4. CAR T cells generated with Next Gen VivoVec particles exhibit increased functional responses *in vitro*



**Left:** anti-CD19 CAR<sup>+</sup> T cells were generated using the indicated VivoVec particles. Anti-CD19 CAR T<sup>+</sup> cells were cultured with Nalm6 tumor cells for 22 hours followed by supernatant cytokine analysis by MSD®. **Right:** anti-CD19 CAR<sup>+</sup> T cells generated with indicated VivoVec particles were serial-stimulated with Nalm6 tumor cells every 2–3 days. Total Nalm6 tumor cells were tracked over time on an IncuCyte®.

## Summary

- Next Gen VivoVec particles incorporating additional T cell costimulatory molecules enhance particle binding and T cell activation
- Next Gen VivoVec particles improve T cell transduction and produce greater numbers of T cells *in vitro*
- CAR T cells generated with Next Gen VivoVec particles exhibit phenotypic properties associated improved persistence
- CAR T cells generated with Next Gen VivoVec particles have greater antigen-specific cytokine production and *in vitro* tumor killing capacity
- Next Gen VivoVec particles generated more CAR<sup>+</sup> T cells in the blood and enhanced antitumor activity *in vivo*, at lower doses than 1<sup>st</sup> Gen particles, potentially enabling lower clinical doses
- See more exciting work from Umoja at Posters **#366** and **#375**