

Abstract

Adoptive cell therapies featuring *ex vivo* expanded autologous T cells engineered to express tumor-targeting chimeric antigen receptors (CAR T cells) have revolutionized the treatment of B cell malignancies, leading to long-term remission in 30-40% of certain patient populations. Despite the promising clinical efficacy of CAR T cells in hematologic malignancies, major limitations hinder their widespread application, including challenges to patient access, complex manufacturing, and high cost. To overcome these challenges, we have developed VivoVec, an engineered lentiviral particle-based platform harboring a CAR transgene that is being developed for off-the-shelf use for the generation of CAR T cells *in vivo*. To achieve specific and efficient *in vivo* transduction, VivoVec particles are pseudotyped with the Cocal fusion glycoprotein and engineered to express T cell binding, activating, and costimulatory ligands. We evaluated multiple novel surface engineering approaches, including incorporation of an anti-CD3 single chain variable fragment combined with a panel of T cell costimulatory ligands, such as CD80, into the particles' surfaces to initiate T cell activation in conjunction with co-stimulation and particle binding to facilitate efficient transduction. CAR T cells generated with VivoVec particles exhibited a less-differentiated, central memory-like phenotype and CAR-antigen specific polyfunctionality, including proliferation and tumor cell killing *in vitro*. Finally, we observed that VivoVec particles generated CAR T-cells *in vivo* with potent antitumor activity in a humanized NSG mouse model of B cell malignancies. Overall, our results suggest that the collective mechanism of action of VivoVec particles to initiate *in vivo* CAR T cell generation and subsequent anti-tumor immune responses is enabled by particle surface-displayed ligands that promote T cell binding, activation, and costimulation, rendering T-cells competent for transduction while optimizing their immunophenotype and function.

Fig 2. Incorporation of additional costimulatory molecules further enhances T cell activation and binding by VivoVec particles

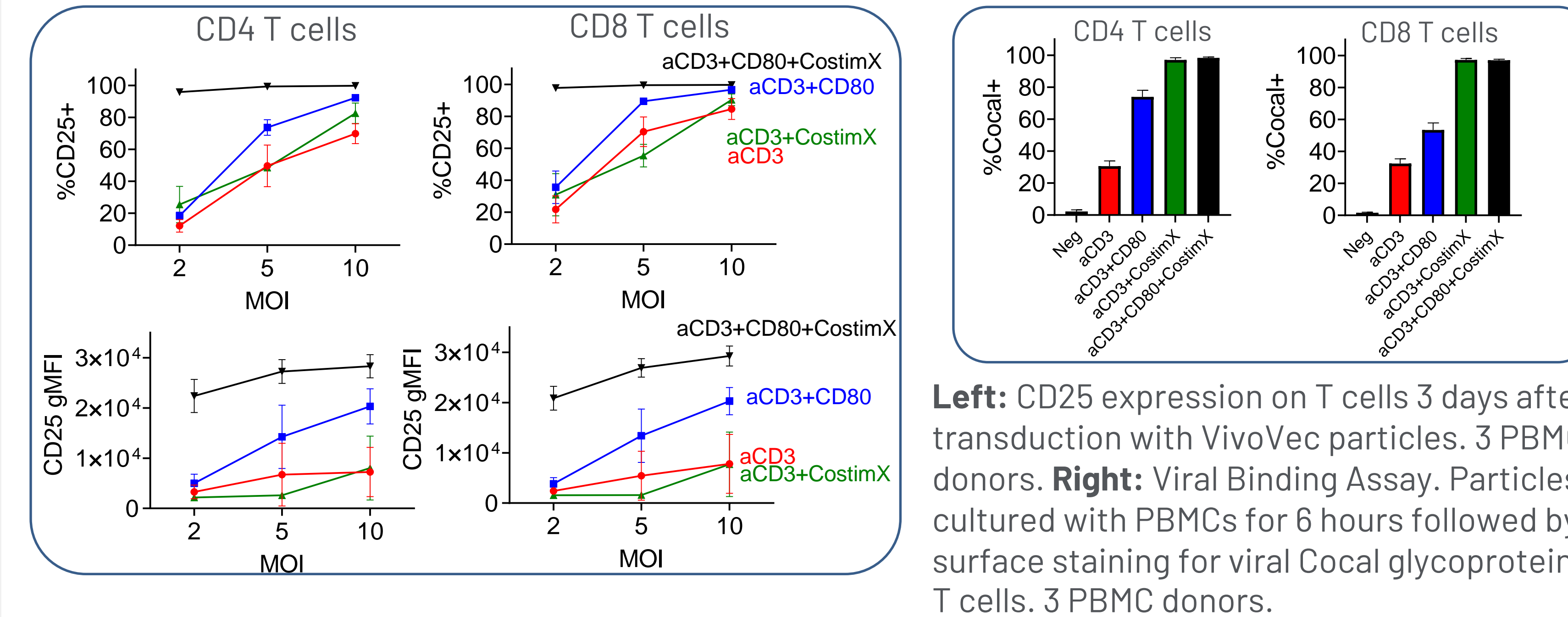
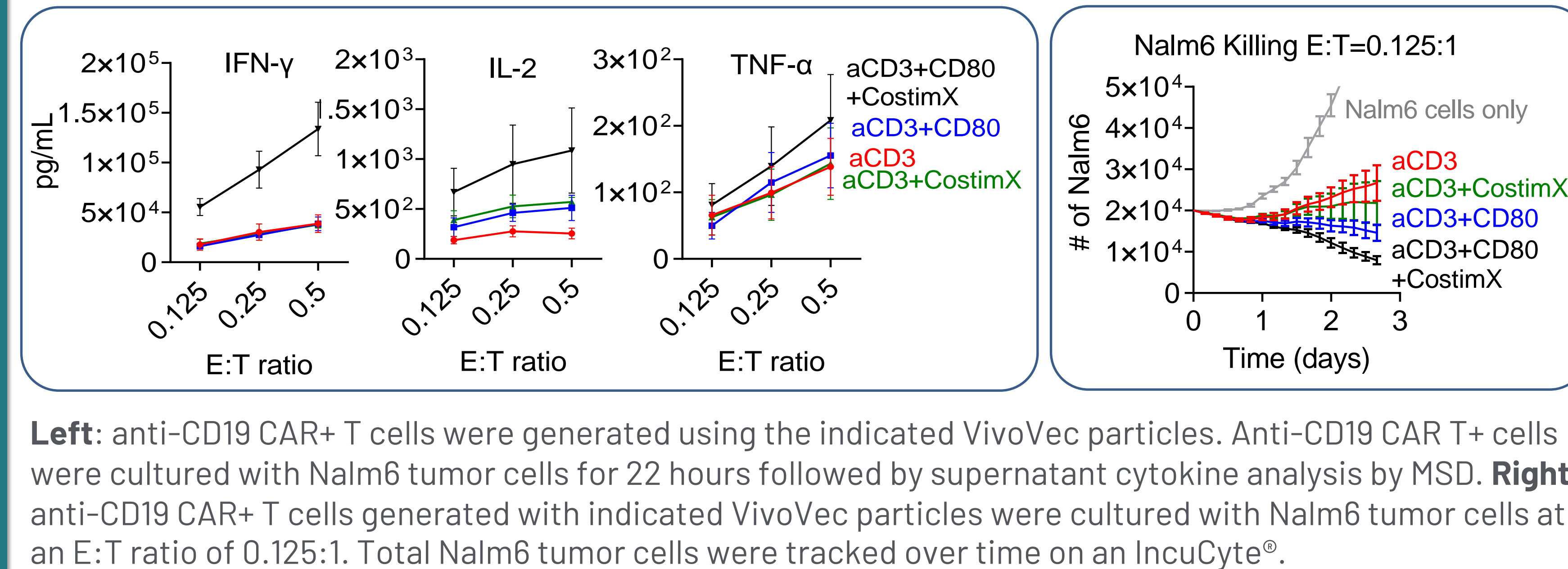
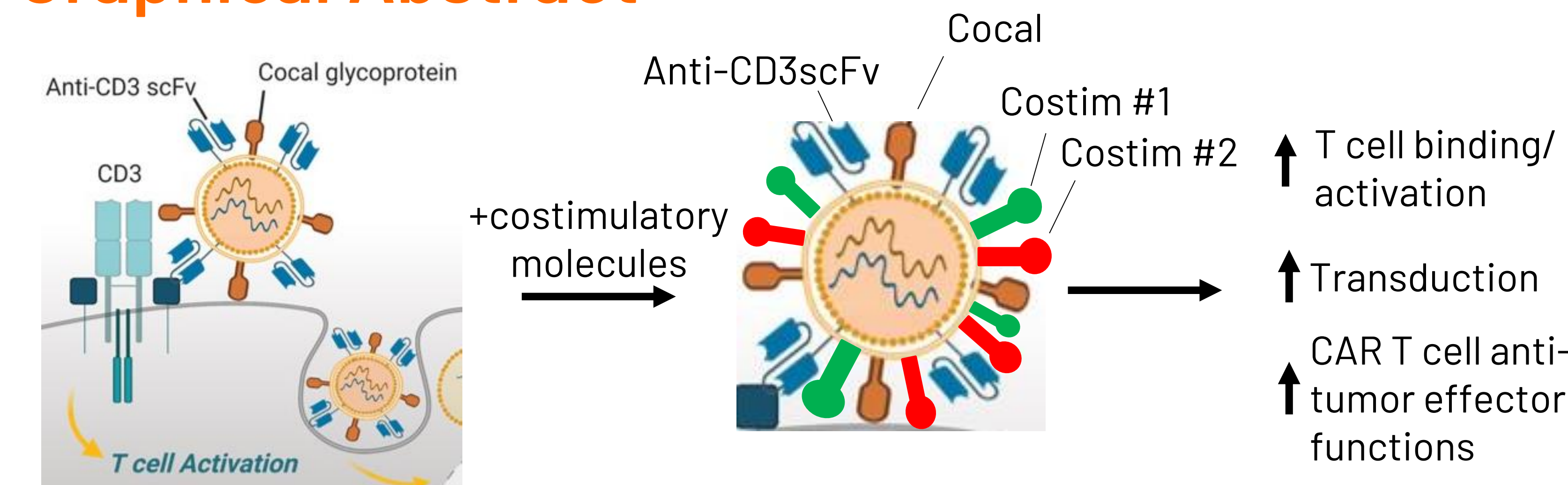


Fig 5. CAR T cells generated with enhanced VivoVec particles exhibit increased cytokine production and cytotoxicity *in vitro*



Graphical Abstract



VivoVec lentiviral vector particles contain an anti-CD3scFv and Cocal glycoprotein to facilitate transduction and delivery of chimeric antigen receptors (CARs) to T cells *in vivo*. VivoVec particles can be further augmented by the addition of T cell costimulatory molecules, resulting in enhanced activation and transduction of T cells and increased anti-tumor effector functions of the resulting CAR-T cells.

Fig 3. Particles incorporating costimulatory molecules increase transduction frequency and T cell expansion

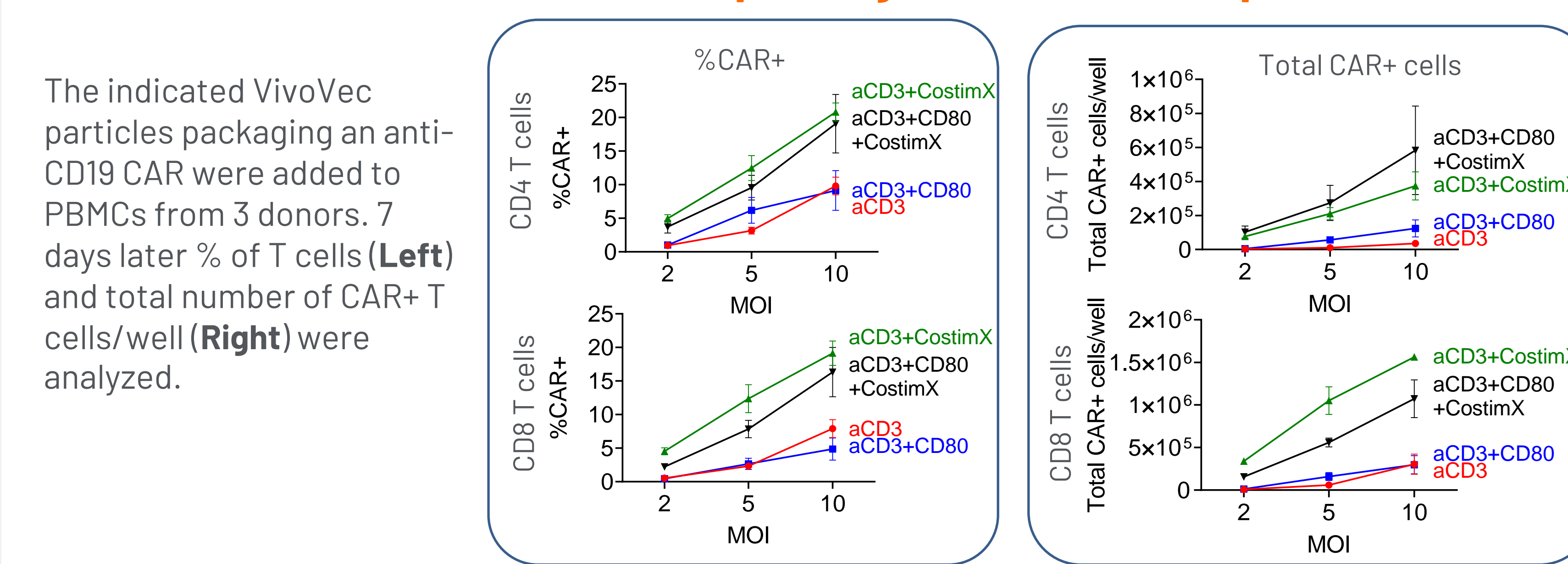


Fig 6. Particles containing costimulatory molecules demonstrate enhanced tumor control in primary tumor response and re-challenge in an *in vivo* tumor xenograft model

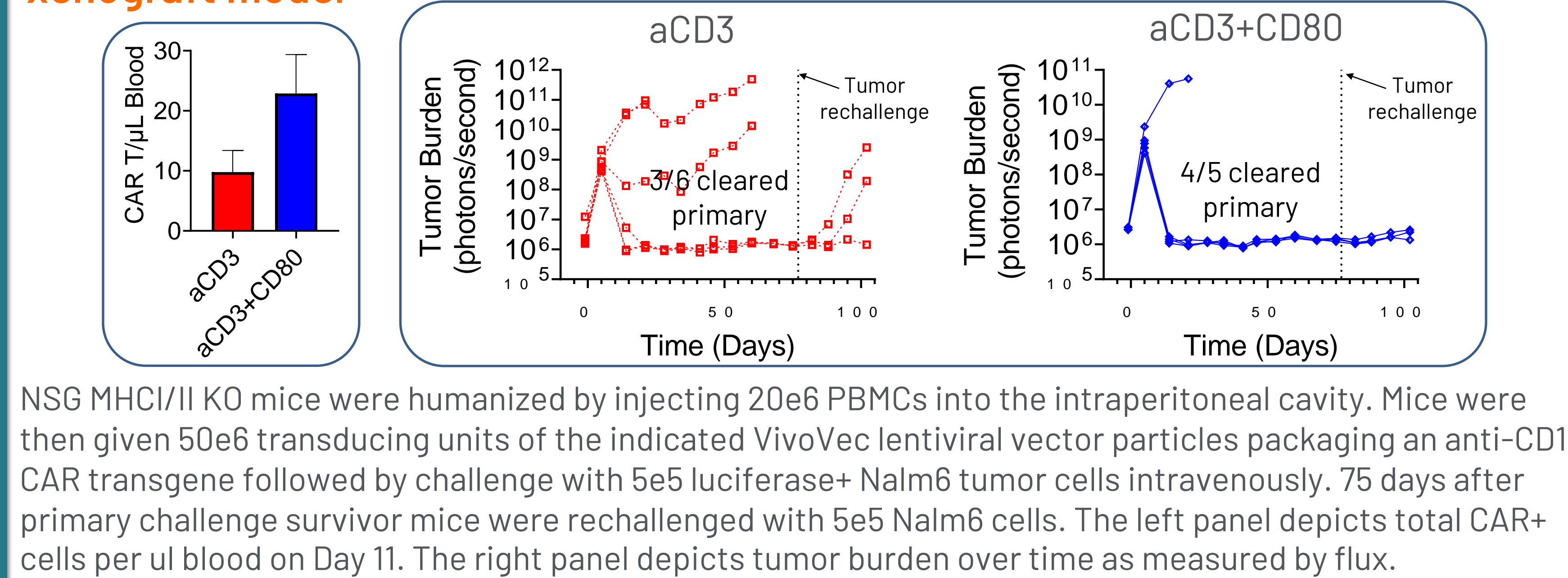


Fig 1. Particles incorporating CD28 costimulatory ligands activate T cells better than particles with CD3scFv alone and generate increased numbers of CAR+ T cells invitro.

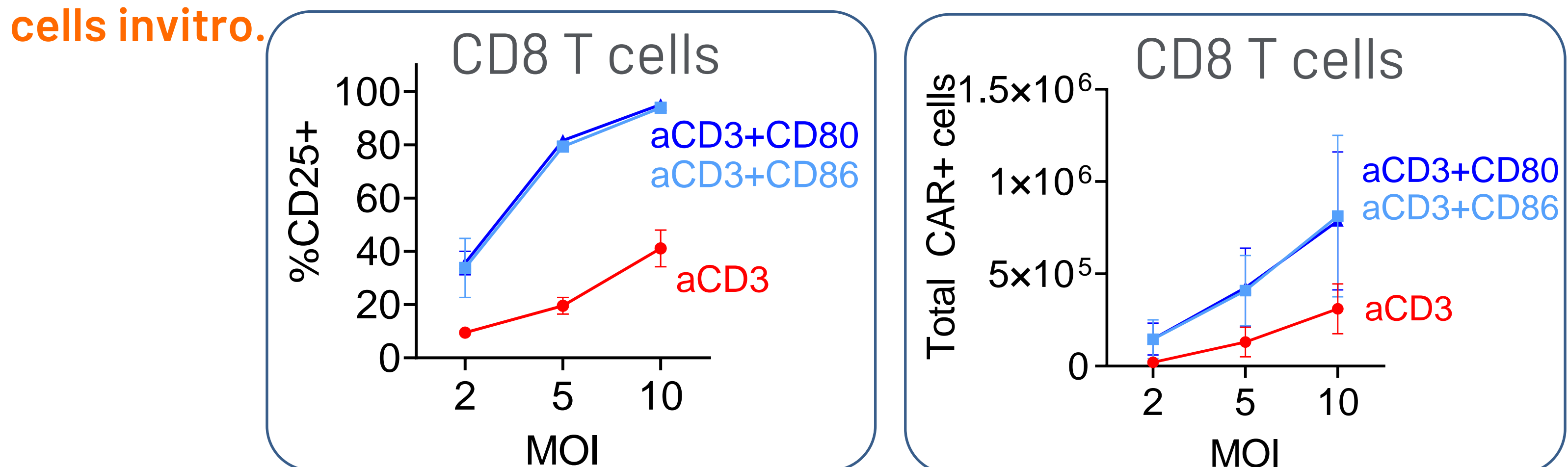
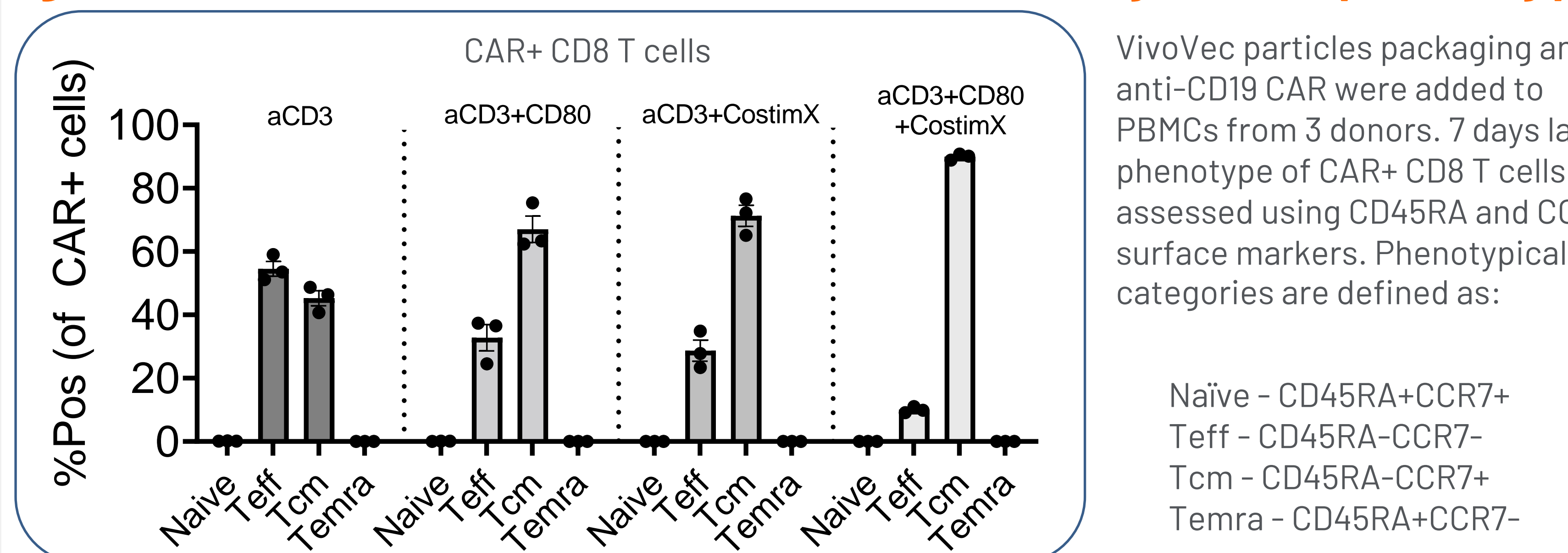


Fig 4. Particles incorporating costimulatory molecules favor generation of T cells with a central memory (Tcm) phenotype



Summary

- VivoVec particles incorporating costimulatory molecules in addition to an aCD3scFv enhance particle binding and activation of T cells
- Enhanced VivoVec particles improve T cell transduction and produce greater numbers of T cells *in vitro*
- CAR T cells generated with enhanced VivoVec particles exhibit a central memory-like phenotype
- CAR T cells generated with enhanced VivoVec particles were more functional in *in vitro* cytokine production and killing assays
- Mice that received enhanced VivoVec particles had greater numbers of CAR+ cells in the blood and exhibited increased tumor clearance - including a robust rechallenge response